

Published in final edited form as:

Zhong Nan Da Xue Xue Bao Yi Xue Ban. 2011 October ; 36(10): 927–938. doi:10.3969/j.issn.1672-7347.2011.10.002.

Multidrug resistance-associated protein 1 (*MRP1/ABCC1*) polymorphism: from discovery to clinical application

YIN Jiye^{1,2} and ZHANG Jianting^{1,2}

¹ Department of Pharmacology/Toxicology, Indiana University School of Medicine, Indianapolis Indiana 46202, USA

² IU Simon Cancer Center, Indiana University School of Medicine, Indianapolis Indiana 46202, USA

Abstract

Multidrug resistance-associated protein 1 (*MRP1/ABCC1*) is the first identified member of ABCC subfamily which belongs to ATP-binding cassette (ABC) transporter superfamily. It is ubiquitously expressed in almost all human tissues and transports a wide spectrum of substrates including drugs, heavy metal anions, toxicants, and conjugates of glutathione, glucuronide and sulfate. With the advance of sequence technology, many *MRP1/ABCC1* polymorphisms have been identified. Accumulating evidences show that some polymorphisms are significantly associated with drug resistance and disease susceptibility. *In vitro* reconstitution studies have also unveiled the mechanism for some polymorphisms. In this review, we present recent advances in understanding the role and mechanism of *MRP1/ABCC1* polymorphisms in drug resistance, toxicity, disease susceptibility and severity, prognosis prediction, and methods to select and predict functional polymorphisms.

Keywords

multidrug resistance-associated protein 1; ABCC1; single nucleotide polymorphism; drug resistance; prognosis; disease susceptibility

Multidrug resistance-associated protein 1 (*MRP1/ABCC1*) is a member of the ATP-binding cassette (ABC) transporter superfamily which contains 49 members in human that are divided into 7 subfamilies, named from ABCA to ABCG (<http://nutrigene.4t.com/humanabc.htm>)^[1-2]. *MRP1/ABCC1* is the first identified gene in the ABCC subfamily and was cloned from a multidrug resistant small cell lung cancer cell line H69AR^[3]. Subsequent studies revealed the important role of *MRP1/ABCC1* as an exporter of drugs and metabolites in many physiological, pathological and pharmacological processes. Thus, polymorphism is likely an important feature of *MRP1/ABCC1* in disease susceptibility, drug response, and treatment outcomes^[4]. In this review, we will evaluate recent advances in discovery of *MRP1/ABCC1* polymorphisms and understanding their potential clinical applications.

1 STRUCTURE AND TISSUE DISTRIBUTION

The *MRP1/ABCC1* gene is located in chromosome 16p13.1 and spans approximately 200 kb. It contains 31 exons and encodes a protein of 1531 amino acid residues with an apparent molecular weight of 180-190 kD^[3-5]. *MRP1/ABCC1* is an atypical ABC transporter with three membrane-spanning domains (MSD) and two cytosolic nucleotide binding domains (NBD)^[6]. While MSD1 and MSD2 each consists of 6 transmembrane (TM) segments, MSD0 has 5 TM segments with a predicted extracellular amino terminus (Fig. 1A). However, recent studies showed that the amino terminus of human MRP1/ABCC1 may have an unusual U-shaped structure which possibly serves as a gate for MRP1/ABCC1 function^[7-9].

The sequence of MSD is highly divergent among different members of ABC transporter family, consistent with MSD's possible function in determining substrate specificity^[10]. Thus, polymorphisms in this domain may affect the substrate spectrum of MRP1/ABCC1. While a typical ABC transporter has two MSDs, the additional MSD0 of human MRP1/ABCC1 is peculiar and its function is not yet fully elucidated. However, our recent studies showed that MSD0 contributes to MRP1/ABCC1 homo-dimerization^[11-12].

In contrast to MSD, NBD is highly conserved among different ABC transporters. It is responsible for binding and hydrolysis of ATP to provide energy for substrate transport^[10]. Similar to other ABC transporters, the NBD of MRP1/ABCC1 has two consensus motifs designated as "Walker A" and "Walker B"^[13] and a third consensus motif designated as ABC-signature motif of approximate 13 amino acids between Walker A and Walker B^[10]. These highly conserved motifs are critical for MRP1/ABCC1 function and a single mutation may abolish the activity of the whole protein^[14-15]. Thus, polymorphisms in NBD may produce inactive MRP1/ABCC1.

MRP1/ABCC1 appears to be ubiquitously expressed in almost all human tissues^[16-18]. Its expression level is high in lung, spleen, testis, kidney, placenta, thyroid, bladder and adrenal gland, but low or no expression in some cells of circulatory system, such as eosinophils, helper T-cells and erythrocytes^[19]. MRP1/ABCC1 is also expressed in blood-brain, blood-testis and blood-cerebrospinal fluid (CSF) barriers, which was thought to contribute to protection of these organs by keeping out toxic substances^[20-21]. Indeed, it has been shown that accumulation of etoposide in CSF increased 10-fold in MRP1/ABCC1 knockout mice^[20]. At the cellular level, in contrast to the apical membrane location of other ABC transporters, MRP1/ABCC1 is predominantly located in the basolateral membrane of polarized cells^[22-23]. Thus, MRP1/ABCC1 likely pumps its substrate into the interstitial space of body, rather than excreting them into bile, urine or gut.

2 SUBSTRATES

MRP1/ABCC1 can transport a wide spectrum of substrates ranging from anticancer drugs to fluorescent dye (Tab. 1). A wide variety of anticancer drugs including anthracyclines, epipodophyllotoxins, vinca alkaloids, camptothecins, methotrexate and mitoxantrone are known substrates of MRP1/ABCC1 and, thus, MRP1/ABCC1 over-expression leads to multidrug resistance in cancer chemotherapy. In addition to anticancer drugs, MRP1/

ABCC1 also transports many other types of drugs, such as anti-HIV drugs. Therefore, *MRP1/ABCC1* gene polymorphisms may affect patient response to chemotherapy of these diseases. Previously, we have shown that G2168A polymorphism significantly reduced *MRP1/ABCC1* activity in resistance to anthracyclines, vinca alkaloids and etoposide^[24].

Another important group of substrates of *MRP1/ABCC1* is organic anion conjugates including glutathione, glucuronides and sulfate conjugates. Transporting these conjugates helps cells to remove toxins and protect tissues from damage^[25-26]. LTC₄, a very important mediator of inflammatory response which controls vascular permeability and smooth muscle contraction, is another high affinity substrate of *MRP1/ABCC1*^[19,27]. Thus, *MRP1/ABCC1* polymorphisms may affect therapeutic efficiency of some LTC₄ targeting drugs, such as montelukast and zileuton^[28-29].

3 POLYMORPHISMS

A large number of naturally occurring *MRP1/ABCC1* polymorphisms have been identified with most studies in Asian and Caucasian populations^[30-37]. A comprehensive list of naturally occurring *MRP1/ABCC1* polymorphisms in different populations can be found in several publicly accessible databases [Pharmacogenetics Research Network: <http://www.pharmgkb.org>; National Central for Biotechnology Information (NCBI): <http://www.ncbi.nlm.nih.gov/snp>; Japanese Single Nucleotide Polymorphisms (JSNP) database: <http://snp.ims.u-tokyo.ac.jp/>; International HapMap Project: www.hapmap.org/].

Most identified *MRP1/ABCC1* polymorphisms are single nucleotide polymorphisms (SNPs), although repeats, insertions and deletions are also found. There are vast ethnical differences in *MRP1/ABCC1* polymorphism distribution and frequency, especially between Asian and Caucasian. For example, G2168A is a common SNP in the Asian population, but it has not been found in Caucasian^[24]. On the contrary, G2012T polymorphism is common in Caucasian, but not found in Asian populations^[31]. Most *MRP1/ABCC1* polymorphisms have a very low frequency (< 5%), which indicating that *MRP1/ABCC1* is a highly conserved gene. The majority of identified polymorphisms are located in the untranslated region (UTR) and introns and few polymorphisms are located in the coding region. Polymorphisms in the coding region are more likely to be functional and can be divided into three types: synonymous (no change in amino acid sequence resulting in a wild-type protein), non-synonymous (change in amino acid sequence resulting in a mutant protein), and nonsense (change to a stop codon resulting in a truncated protein). Up to date, only 14 non-synonymous polymorphisms have been identified with very low frequencies and no nonsense polymorphism has been found (Fig. 1). These non-synonymous polymorphisms were intensively studied both *in vitro* and *in vivo* since they could be easily recreated using site-directed mutagenesis and they might affect the expression and function of *MRP1/ABCC1*^[24-39]. Although the polymorphisms in the non-coding region do not affect the sequence of the protein, they are also important and can be used as genetic markers^[40-41].

4 ASSOCIATION OF *MRP1/ABCC1* POLYMORPHISMS WITH THERAPEUTIC RESPONSE

As discussed above, many therapeutic drugs are substrates of *MRP1/ABCC1*. Thus, it is conceivable that some *MRP1/ABCC1* polymorphisms may affect treatment responses and toxicities. Tab. 2 lists *MRP1/ABCC1* polymorphisms that have been studied for their association with therapeutic responses. One of these polymorphisms, G2012T which was first identified by Conrad et al.^[34] in Caucasian population, has been extensively studied. It causes mutation of a highly conserved Gly⁶⁷¹ to Val. Investigation of its potential relationship with response to atorvastatin in treatment of hypercholesterolemia, telatinib in treatment of solid tumors, and induction therapy of leukemia, however, showed no significant correlation with treatment responses^[42-44]. Consistent with these clinical observations, *in vitro* studies also showed that the mutant *MRP1/ABCC1* carrying this mutation had no detectable difference in drug transport activity from the wild type *MRP1/ABCC1*^[34]. Thus, the G2012T polymorphism may not have functional impact on chemotherapy.

Another extensively studied polymorphism is G4002A, a synonymous SNP located in exon 28. Several studies exploring the correlation of G4002A polymorphism and responses to anticancer drugs gemcitabine, cisplatin, taxanes and methotrexate showed no significant association in pancreatic cancer patients^[45-47]. However, Lee et al.^[48] found that this polymorphism was strongly associated with the response of patients with major depressive disorder to antidepressant citalopram. Although patients with the G4002A polymorphism had a 4.7-fold increase in citalopram response, there is no evidence that G4002A polymorphism of *MRP1/ABCC1* in the blood-brain barrier affects citalopram uptake and if citalopram is a substrate of *MRP1/ABCC1*. Another non-synonymous polymorphism located in exon 28, A4009G, was found to correlate with methotrexate therapeutic efficacy in a study of 374 chronic plaque psoriasis patients who received methotrexate monotherapy^[49]. It was found that the heterozygous A4009G in the responders is significantly higher than that in non-responders, suggesting that the A4009G polymorphism may increase methotrexate responses. However, it has not yet been determined if the A4009G polymorphism affects *MRP1/ABCC1* expression, trafficking, or function. Future studies on the possible effects of the A4009G polymorphism on these aspects of *MRP1/ABCC1* are needed.

A well studied polymorphism that has been shown to significantly reduce drug transport activity of *MRP1/ABCC1* is G2168A^[24]. It has also been shown to increase chemotherapy response in advanced ovarian cancer patients^[50]. In the study of advanced ovarian cancer patients, several other polymorphisms of *MRP1/ABCC1* (T825C, T1062C, T1684C, C2007T and G4002A) were also investigated. However, none of these polymorphisms were found to significantly associate with chemotherapy responses. Thus, the G2168A polymorphism may be an indicator of chemotherapy response of advanced ovarian cancers. However, whether this polymorphism also affects chemotherapy responses of other human cancers need to be investigated.

In addition to the polymorphisms in the coding region, some polymorphisms in the non-coding region of *MRP1/ABCC1* are also found to associate with drug responses. Two such polymorphisms in the non-coding region are IVS23 G-1960A and IVS9 T-176C located in intron 23 and 9, respectively. They both have been shown to significantly associate with methotrexate response in psoriasis patients and patients carrying these polymorphisms appear to have worse response to methotrexate treatment^[49]. Another example of polymorphisms in the non-coding region is IVSI C-14840T which is located in intron 1 and has been found to correlate with significantly higher response to both montelukast and zileuton in asthma patients than wild-type homozygotes^[28-29]. Thus, polymorphisms in *MRP1/ABCC1* may affect montelukast and zileuton response and lung function. Interestingly, in another study of two independent cohorts, polymorphisms of *MRP1/ABCC1* in the 3'-UTR (G3361A and A2615G) and IVS14 C-1575T also significantly correlate with lung function^[51]. While 3'-UTR G3361A correlates with higher forced expiratory volume at one second (FEV1), 3'-UTR A2615G correlates with lower FEV1. Another polymorphism, IVS14 C-1575T in the intron 14 of *MRP1/ABCC1*, correlates with highly excessive FEV1 decline. However, how these polymorphisms in the non-coding region possibly affect *MRP1/ABCC1* is not yet known. It is also unknown if *MRP1/ABCC1* plays any role in lung function. While the polymorphisms in the UTR may affect the translation and expression of *MRP1/ABCC1*, the polymorphisms in the intron may affect RNA processing. Clearly, these hypothetical mechanisms of action and the role of *MRP1/ABCC1* in lung function needs to be investigated in the near future.

5 ASSOCIATION OF *MRP1/ABCC1* POLYMORPHISMS WITH PROGNOSIS PREDICTION

Based on the above discussion of association of *MRP1/ABCC1* polymorphisms with therapeutic response, it is tempting to speculate that polymorphisms of *MRP1/ABCC1* may be used as markers to predict prognosis. Indeed, two polymorphisms have been shown to associate with prognosis (Tab. 3). In a study of possible contribution of four non-synonymous polymorphisms of *MRP1/ABCC1* to neuroblastoma outcome in a cohort of 195 Caucasian patients, it was found that the presence of the G2010T polymorphism has significant improvement in outcome^[52]. It was also found that the G2010T polymorphism reduces the stability and expression level of *MRP1/ABCC1* mRNA. Hence, it is possible that patients with the G2010T polymorphism may have reduced level of *MRP1/ABCC1*, which would enhance drug response and increase chemotherapy efficacy. In another study of correlating 5'-UTR G-1666A polymorphism with hepatocellular carcinoma (HCC) outcome in 162 Chinese patients, it was found that the mutant genotype carriers had better prognosis with increased 4-year disease free survival^[53]. Using *in vitro* electrophoretic mobility shift assay (EMSA), these authors also found that the mutant allele had much less binding affinity to nuclear proteins, suggesting that this promoter polymorphism may cause decreased transcription of *MRP1/ABCC1*. However, whether this promoter polymorphism inhibits *MRP1/ABCC1* transcription has not yet been demonstrated. It is also unknown if the nuclear proteins that bind to this region are involved in the transcription of *MRP1/ABCC1*. Nevertheless, these polymorphisms may be used as makers predicting prognosis and survival in neuroblastoma and HCC.

6 ASSOCIATION OF *MRP1/ABCC1* POLYMORPHISMS WITH DRUG TOXICITY

Since some toxicants and drug metabolites are also substrates of *MRP1/ABCC1*, possible association of *MRP1/ABCC1* polymorphisms and drug toxicity is also of importance and interest to investigate. In this regard, correlation of *MRP1/ABCC1* polymorphisms and drug-induced neuropathy is mostly studied (Tab. 4). In a recent study correlating polymorphisms of *MRP1/ABCC1* (IVS9A8G, IVS11C-48T, T1684C, IVS18C-30G, G4002A and IVS30A18G) with irinotecan-induced neutropenia in cancer patients, it was found that the TT genotype carriers of IVS11 C-48T had significant lower neutrophil count (ANC) in patients receiving irinotecan monotherapy^[54]. Irinotecan-induced neutropenia is thought to be due to production of the cytotoxic irinotecan metabolite, SN-38, which is a substrate of *MRP1/ABCC1*. Consistent with this study, *MRP1/ABCC1* polymorphism has also been found to correlate with peripheral neuropathy induced by vincristine^[55]. In this study of 833 myeloma patients, it was found that the carriers of *MRP1/ABCC1* polymorphism IVS16 A1695T were more likely to develop vincristine-induced peripheral neuropathy than the wild type carriers. Similar to SN-38, it is also speculated that this polymorphism may decrease *MRP1/ABCC1*-mediated transport of vincristine and, thus, increases vincristine-induced peripheral neuropathy. However, the molecular mechanisms need further investigation.

One interesting polymorphism is G2012T, which shows correlation with doxorubicin toxicity in non-Hodgkin lymphoma patients^[56]. The patients with this polymorphism have more anthracycline-induced cardiotoxicity than the wild-type patients. It was thought that the special subcellular localization of *MRP1/ABCC1* in cardiomyocytes, in both plasma and lysosome membranes, permits sequestration of doxorubicin in lysosomes and prevent doxorubicin cardiotoxicity^[17-57]. However, it has been demonstrated previously that the G2012T polymorphism of *MRP1/ABCC1* has no effect on its function and substrate transport activity^[34]. Thus, it is not clear how this polymorphism affects anthracycline-induced cardiotoxicity. Furthermore, since multidrug chemotherapy was used for these cohorts of patients, interpretation of these observations should be cautious. Doxorubicin mono-therapy and further investigation of G2012T mutation on *MRP1/ABCC1* activity in transporting doxorubicin would help clarify this issue.

Several other polymorphisms of *MRP1/ABCC1* (IVS3 G-3198A, IVS4 G409A, IVS5G413A, IVS5 A-7942G, IVS5G-1641A and IVS23 G-1960A) have been found to significantly correlate with methotrexate toxicity in liver and GI tract of psoriasis patients^[49]. All these polymorphisms are located in introns and form a haplotype although it is not yet known if they affect the expression of *MRP1/ABCC1* individually or as a haplotype. Based on the above discussion, *MRP1/ABCC1* polymorphisms are likely important genetic indicators in drug toxicity during chemotherapy.

7 ASSOCIATION OF *MRP1/ABCC1* POLYMORPHISMS WITH DISEASE SUSCEPTIBILITY AND SEVERITY

Association of *MRP1/ABCC1* with disease susceptibility has also been identified (Tab. 5). In a case control study of 500 lung cancer patients and 517 cancer free control subjects in Chinese population, Wang et al. [58] detected the association of three polymorphisms in the 3'-UTR of *MRP1/ABCC1* (C543T, T866A and T1512C) with lung cancer susceptibility. They found that subjects carrying mutant allele of 3'-UTR T866A had an increased risk of lung cancer. However, the other two polymorphisms had no significant correlation with lung cancer susceptibility. Further investigation showed that these three polymorphisms form a haplotype and the GTA haplotype was associated with increased risk of lung cancer compared with the most prevailing AAA haplotype. Therefore, this polymorphism haplotype may increase lung cancer predisposition in Chinese population. We recently identified association of another non-synonymous polymorphism G2168A with lung cancer susceptibility[59]. In our study of 77 lung cancer patients and 71 control individuals in Chinese population, we showed that the subjects carrying the G2168A allele had 3.5 fold increased risk (adjusted OR = 3.42; 95% CI, 1.29 - 9.06; $P=0.013$) of lung cancer compared with wild-type carriers. Further stratified analysis showed that the elderly people (> 50 years) carrying mutant allele of this polymorphisms were more likely to develop lung cancer (adjusted OR, 4.10; 95% CI, 1.25-13.48; $P=0.020$) than younger ones. Taken together, it is possible that *MRP1/ABCC1* polymorphisms may play important roles in lung cancer susceptibility. Although the mechanism of *MRP1/ABCC1* action in lung cancer susceptibility is unknown, it is tempting to speculate that *MRP1/ABCC1* may protect lung tissues against carcinogens by preventing them from entering bronchial epithelial cells. Carriers of these *MRP1/ABCC1* polymorphisms are likely more susceptible to carcinogenesis due to reduced protection by *MRP1/ABCC1*. This possibility is consistent with our observation that the G2168A polymorphism decreases *MRP1/ABCC1* function in drug transport activity (unpublished observations). However, further clinical studies are needed to test this possibility.

Possible impact of *MRP1/ABCC1* polymorphisms on disease severity has also been reported in other studies (Tab. 5). In a study of 203 cystic fibrosis (CF) patients, it was found that the G-260C polymorphism in the 5'-UTR of *MRP1/ABCC1* significantly increased CF severity[60]. Patients with CC genotype had earlier onset of chronic colonization by *Pseudomonas aeruginosa* (PA). Although *in vitro* study showed no impact of this polymorphism on promoter transcriptional activity, mRNA levels, basal and cAMP-induced anion transport, the possibility that this polymorphism affects translation/synthesis of *MRP1/ABCC1* and, thus, its expression level cannot be ruled out.

In another study of five *MRP1/ABCC1* polymorphisms (3'-UTR T866A, 3'-UTR G3361A, 5'-UTR C-435G, IVS1 T5977G and IVS14 C-1575T) and their possible effect on chronic obstructive pulmonary disease (COPD) severity, it was found that the 3'-UTR T866A was associated with higher FEV1 level and less airway wall inflammation while the 3'-UTR G3361A was associated with lower FEV1 level and higher inflammation. However, the other three polymorphisms have no significant association with COPD severity[61]. The

mechanism of the 3'-UTR T866A in affecting COPD severity remains unknown. However, it is speculated that 3'-UTR T866A may affect *MRP1/ABCC1* mRNA stability together with another 3'-UTR polymorphism 801 C > GR^[51-61] They were found to be in complete linkage disequilibrium^[40]. Clearly, *MRP1/ABCC1* polymorphisms are likely associated with lung cancer susceptibility and with COPD and CF disease severity. However, whether and how each polymorphism possibly affects disease susceptibility and severity need to be investigated in the future.

8 CONCLUSIONS SPECTIVES

Since the discovery of *MRP1/ABCC1* in 1992, many *MRP1/ABCC1* polymorphisms have been identified. Most of the identified polymorphisms are synonymous and have low frequency, indicating that *MRP1/ABCC1* is a highly conserved gene. Some of the *MRP1/ABCC1* polymorphisms have been found to associate with drug response, prognosis, toxicity, disease susceptibility and severity. Some of these polymorphisms have also been shown to affect *MRP1/ABCC1* expression or function which may indicate the underling mechanism of association with the observed phenotype. With the advances of next generation sequencing, International HapMap Project and 1 000 Genomes Project^[62-63], more *MRP1/ABCC1* polymorphisms are likely to be identified. However, identifying functional *MRP1/ABCC1* polymorphisms and their mechanisms of action will not be easy. Thus, both opportunities and challenges exist.

Because not every polymorphism is functional, selecting potentially functional polymorphisms for further clinical relevance study is important considering the large number of polymorphisms is to be identified. Use of *in silico* and bioinformatics tools such as SIFT, PANTHER and Polyphen algorithms to detect sequence conservation can help identify the likely functional polymorphism since sequences that are highly conserved across different species tend to be functionally important^[64-66]. However, this strategy should be used with caution due to both false positive and negative predictions. For example, G689A, G1057A and G3173A polymorphisms of *MRP1/ABCC1* are predicted as deleterious polymorphisms using SIFT. However, none of these polymorphisms adversely affects *MRP1/ABCC1* function^[24-39].

Examination of polymorphism databases shows that most polymorphisms are located in introns and UTRs. In addition, some polymorphisms located in the exons are synonymous polymorphisms. Thus, study of sequence conservation will unlikely be able to predict if these polymorphisms are functional. For these polymorphisms, a genome-wide approach to identify polymorphisms of positive and negative selection is helpful^[41-67]. Positive selection is an evolutionary process and the positively selected polymorphisms contribute to the favorable phenotype of species and, thus, these polymorphisms may be of higher frequency in the population and important for the gene function^[66-68]. Opposite to positive selection, negative selection is the decline of disadvantage phenotype and harmful and, thus, the negatively selected polymorphisms usually have very low frequency (minor allele frequency < 0. 05) in the population although they may be important for the function and rare drug adverse effects^[66]. Both strategies have been used to identify functional *MRP1/ABCC1* polymorphisms^[38,41,56,67]. However, it is noteworthy that combination of sequence

conservation and evolutionary features may be more powerful than any approach alone to predict and identify functional polymorphisms.

Another challenge is to understand how each polymorphism affects gene function. While it is easy to study the effect of the non-synonymous polymorphisms on the structure and function of MRP1/ABCC1 by re-creating the mutant protein and analyzing the protein in cell lines^[24-39], it is challenging to investigate the synonymous or non-coding region polymorphisms due to complexity of their functional gene effect by different mechanisms such as transcription, splicing, RNA stability, and combined haplotype^[40, 49, 53, 69-70].

Acknowledgments

Foundation items This work was supported by a grant from NIH (ROI CA120221) and China Scholarship Council and Scholarship Award for Excellent Doctoral Student granted by Ministry of Education of China.

Biography

YIN Jiye, Ph.D., mainly engaged in the research of pharmacogenetics.

REFERENCES

- [1]. Choudhuri S, Klaassen CD. Structure, function, expression, genomic organization, and single nucleotide polymorphisms of human ABCB1 (MDR1), ABCC (MRP), and ABCG2 (BCRP) efflux transporters[J]. *Int J Toxicol*. 2006; 25(4):231–259. [PubMed: 16815813]
- [2]. Mo W, Zhang JT. Oligomerization of human ATP-binding cassette transporters and its potential significance in human disease [J]. *Expert Opin Drug Metab Toxicol*. 2009; 5(9):1049–1063. [PubMed: 19637987]
- [3]. Cole SP, Bhardwaj G, Gerlach JH, et al. Overexpression of a transporter gene in a multidrug-resistant human lung cancer cell line[J]. *Science*. 1992; 258(5088):1650–1654. [PubMed: 1360704]
- [4]. Conseil G, Deeley RG, Cole SP. Polymorphisms of MRP1(ABCC1) and related ATP-dependent drug transporters [J]. *Pharmacogenet Genomics*. 2005; 15(8):523–533. [PubMed: 16006996]
- [5]. Grant CE, Kurz EU, Cole SP, et al. Analysis of the intron-exon organization of the human multidrug-resistance protein gene (MRP) and alternative splicing of its mRNA [J]. *Genomics*. 1997; 45(2):368–378. [PubMed: 9344662]
- [6]. Mo, W.; Liu, JY.; Zhang, JT. Biochemistry and pharmacology of human ABCC1/MRP1 and its role in detoxification and in multidrug resistance of cancer chemotherapy in recent advances on cancer research and therapy[M]. In: Pestka, S.; Shi, Y.; Liu, XY., editors. *Recent advances on cancer research and therapy*. Elsevier; 2011. in press
- [7]. Yang Y, Chen Q, Zhang JT. Structural and functional consequences of mutating cysteine residues in the amino terminus of human multidrug resistance-associated protein 1 [J]. *J Biol Chem*. 2002; 277(46):44268–44277. [PubMed: 12235150]
- [8]. Chen Q, Yang Y, Li L, et al. The amino terminus of the human multidrug resistance transporter ABCC1 has a U-shaped folding with a gating function [J]. *J Biol Chem*. 2006; 281(41):31152–31163. [PubMed: 16914551]
- [9]. Chen Q, Yang Y, Liu Y, et al. Cytoplasmic retraction of the amino terminus of human multidrug resistance protein 1 [J]. *Biochemistry*. 2002; 41(29):9052–9062. [PubMed: 12119019]
- [10]. Hipfner DR, Deeley RG, Cole SP, et al. Structural, mechanistic and clinical aspects of MRP1 [J]. *Biochim biophys Acta*. 1999; 1461(2):359–376. [PubMed: 10581367]
- [11]. Yang Y, Mo W, Zhang JT. Role of transmembrane segment 5 and extracellular loop 3 in the homodimerization of human AB-CC1[J]. *Biochemistry*. 2010; 49(51):10854–10861. [PubMed: 21090806]

- [12]. Yang Y, Liu Y, Dong Z, et al. Regulation of function by dimerization through the amino-terminal membrane-spanning domain of human ABCC1/MRP1 [J]. J Biol Chem. 2007; 282(12):8821–8830. [PubMed: 17264072]
- [13]. Walker JE, Saraste M, Runswick MJ, et al. Distantly related sequences in the alpha-and beta-subunits of ATP synthase, myosin, kinases and other ATP-requiring enzymes and a common nucleotide binding fold[J]. EMBO J. 1982; 1(8):945–951. [PubMed: 6329717]
- [14]. Szentpétery Z, Sarkadi B, Bakos E, et al. Functional studies on the MRP1 multidrug transporter: characterization of ABC-signature mutant variants[J]. Anticancer Res. 2004; 24(2A):449–455. [PubMed: 15152943]
- [15]. Szentpétery Z, Kern A, Liliom K, et al. The role of the conserved glycines of ATP-binding cassette signature motifs of MRP1 in the communication between the substrate-binding site and the catalytic centers[J]. J Biol Chem. 2004; 279(40):41670–41678. [PubMed: 15252017]
- [16]. Zaman GJ, Versantvoort CH, Smit JJ, et al. Analysis of the expression of MRP, the gene for a new putative transmembrane drug transporter, in human multidrug resistant lung cancer cell lines[J]. Cancer Res. 1993; 53(8):1747–1750. [PubMed: 8467491]
- [17]. Flens MJ, Zaman GJ, Van der Valk P, et al. Tissue distribution of the multidrug resistance protein [J]. Am J Pathol. 1996; 148(4):1237–1247. [PubMed: 8644864]
- [18]. St-Pierre MV, Serrano MA, Macias RI, et al. Expression of members of the multidrug resistance protein family in human term placenta [J]. Am J Physiol Regul Integr Comp Physiol. 2000; 279(4):R1495–R1503. [PubMed: 11004020]
- [19]. Chang XB. A molecular understanding of ATP-dependent solute transport by multidrug resistance-associated protein MRP1 [J]. Cancer Metastasis Rev. 2007; 26(1):15–37. [PubMed: 17295059]
- [20]. Wijnholds J, deLange EC, Scheffer GL, et al. Multidrug resistance protein 1 protects the choroid plexus epithelium and contributes to the blood-cerebrospinal fluid barrier [J]. J Clin Invest. 2000; 105(3):279–285. [PubMed: 10675353]
- [21]. Mercier C, Masseguin C, Roux F, et al. Expression of P-glycoprotein(ABCB1) and Mrp1(ABCC1) in adult rat brain: focus on astrocytes[J]. Brain Res. 2004; 1021(1):32–40. [PubMed: 15328029]
- [22]. Evers R, Zaman GJ, van Deemter L, et al. Basolateral localization and export activity of the human multidrug resistance-associated protein in polarized pig kidney cells[J]. J Clin Invest. 1996; 97(5):1211–1218. [PubMed: 8636432]
- [23]. Roelofsen H, Vos TA, Schippers IJ, et al. Increased levels of the multidrug resistance protein in lateral membranes of proliferating hepatocyte-derived cells [J]. Gastroenterology. 1997; 112(2): 511–521. [PubMed: 9024305]
- [24]. Yin JY, Huang Q, Yang Y, et al. Characterization and analyses of multidrug resistance-associated protein 1 (*MRP1/ABCC1*) polymorphisms in Chinese population[J]. Pharmacogenet Genomics. 2009; 19(3):206–216. [PubMed: 19214144]
- [25]. Leslie EM, Deeley RG, Cole CP. Toxicological relevance of the multidrug resistance protein 1, MRP1 (ABCC1) and related transporters[J]. Toxicology. 2001; 167(1):3–23. [PubMed: 11557126]
- [26]. Suzuki H, Sugiyama Y. Excretion of GSSG and glutathione conjugates mediated by MRP1 and cmoat/MRP2 [J]. Semin Liver Dis. 1998; 18(4):359–376. [PubMed: 9875554]
- [27]. Leier I, Jedlitschky G, Buchholz U, et al. Characterization of the ATP-dependent leukotriene C4 export carrier in mastocytoma cells[J]. Eur J Biochem. 1994; 220(2):599–606. [PubMed: 8125120]
- [28]. Lima JJ, Zhang S, Grant A, et al. Influence of leukotriene pathway polymorphisms on response to montelukast in asthma [J]. Am J Respir Crit Care Med. 2006; 173(4):379–385. [PubMed: 16293801]
- [29]. Tantisira KG, Lima J, Sylvia J, et al. 5-lipoxygenase pharmacogenetics in asthma: overlap with cys-leukotriene receptor antagonist loci [J]. Pharmacogenet Genomics. 2009; 19(3):244–247. [PubMed: 19214143]

- [30]. Saito S, Iida A, Sekine A, et al. Identification of 779 genetic variations in eight genes encoding members of the ATP-binding cassette, subfamily C (ABCC/MRP/CFTR) [J]. *J Hum Genet.* 2002; 47(4):147–171. [PubMed: 12166651]
- [31]. Moriya Y, Nakamura T, Horinouchi M, et al. Effects of polymorphisms of MDR1, MRP1, and MRP2 genes on their mRNA expression levels in duodenal enterocytes of healthy japanese subjects[J]. *Biol Pharm Bull.* 2002; 25(10):1356–1359. [PubMed: 12392094]
- [32]. Oselin K, Mrozikiewicz PM, Gaikovitch E, et al. Frequency of MRP1 genetic polymorphisms and their functional significance in caucasians: detection of a novel mutation G816a in the human MRP1 gene[J]. *Eur J Clin Pharmacol.* 2003; 59(4):347–350. [PubMed: 12856092]
- [33]. Fukushima-Uesaka H, Saito Y, Tohkin M, et al. Genetic variations and haplotype structures of the ABC transporter gene AB-CC1 in a japanese population[J]. *Drug Metab Pharmacokinet.* 2007; 22(1):48–60. [PubMed: 17329911]
- [34]. Conrad S, Kauffmann HM, Ito K, et al. Identification of human multidrug resistance protein 1(MRP1) mutations and characterization of a G671V substitution[J]. *J Hum Genet.* 2001; 46(11): 656–663. [PubMed: 11721885]
- [35]. Perdu J, Germain DP. Identification of novel polymorphisms in the pM5 and MRP1(ABCC1) genes at locus 16p13. 1 and exclusion of both genes as responsible for pseudoxanthoma elasticum[J]. *Hum Mutat.* 2001; 17(1):74–75.
- [36]. Wang H, Hao B, Zhou K, et al. Linkage disequilibrium and haplotype architecture for two ABC transporter genes (ABCC1 and ABCG2) in Chinese population: implications for pharmacogenomic association studies[J]. *Ann Hum Genet.* 2004; 68:563–573. Pt 6. [PubMed: 15598215]
- [37]. Ito S, Ieiri I, Tanabe M, et al. Polymorphism of the ABC transporter genes, MDR1, MRP1 and MRP2/cmoat, in healthy japanese subjects[J]. *Pharmacogenetics.* 2001; 11(2):175–184. [PubMed: 11266082]
- [38]. Wang Z, Sew PH, Ambrose H, et al. Nucleotide sequence analyses of the MRP1 gene in four populations suggest negative selection on its coding region [J]. *BMC Genomics.* 2006; 7:111. [PubMed: 16684361]
- [39]. Létourneau IJ, Deeley RG, Cole SP. Functional characterization of non-synonymous single nucleotide polymorphisms in the gene encoding human multidrug resistance protein 1 (*MRP1/ABCC1*) [J]. *Pharmacogenet Genomics.* 2005; 15(9):647–657. [PubMed: 16041243]
- [40]. Leschziner G, Zabaneh D, Pirmohamed M, et al. Exon sequencing and high resolution haplotype analysis of ABC transporter genes implicated in drug resistance[J]. *Pharmacogenet Genomics.* 2006; 16(6):439–450. [PubMed: 16708052]
- [41]. Wang Z, Wang B, Tang K, et al. A functional polymorphism within the MRP1 gene locus identified through its genomic signature of positive selection [J]. *Hum Mol Genet.* 2005; 14(14): 2075–2087. [PubMed: 15944197]
- [42]. Moretti IR, Cristina AR, Sorkin SA, et al. ABCB1 and ABCC1 expression in peripheral mononuclear cells is influenced by gene polymorphisms and atorvastatin treatment [J]. *Biochem Pharmacol.* 2009; 77(1):66–75. [PubMed: 18851956]
- [43]. Mahjoubi F, Akbari S, Montazeri M, et al. MRP1 polymorphisms(T2684C, C2007T, C2012T, and C2665T) are not associated with multidrug resistance in leukemic patients[J]. *Genet Mol Res.* 2008; 7(4):1369–1374. [PubMed: 19065772]
- [44]. Steeghs N, Gelderblom H, Wessels J, et al. Pharmacogenetics of telatinib, a VEGFR-2 and VEGFR-3 tyrosine kinase inhibitor, used in patients with solid tumors[J]. *Invest New Drugs.* 2011; 29(1):137–143. [PubMed: 19924384]
- [45]. Tanaka M, Okazaki T, Suzuki H, et al. Association of multi-drug resistance gene polymorphisms with pancreatic cancer outcome[J]. *Cancer.* 2011; 117(4):744–751. [PubMed: 20922799]
- [46]. Ranganathan P, Culverhouse R, Marsh S, et al. Methotrexate (MTX) pathway gene polymorphisms and their effects on MTX toxicity in caucasian and african american patients with rheumatoid arthritis[J]. *J Rheumatol.* 2008; 35(4):572–579. [PubMed: 18381794]
- [47]. Marsh S, Paul J, King CR, et al. Pharmacogenetic assessment of toxicity and outcome after platinum plus taxane chemotherapy in ovarian cancer: the scottish randomised trial in ovarian cancer [J]. *J Clin Oncol.* 2007; 25(29):4528–4535. [PubMed: 17925548]

- [48]. Lee SH, Lee MS, Lee JH, et al. MRP1 polymorphisms associated with citalopram response in patients with major depression[J]. *J Clin Psychopharmacol.* 2010; 30(2):116–125. [PubMed: 20520284]
- [49]. Warren RB, Smith RL, Campalani E, et al. Genetic variation in efflux transporters influences outcome to methotrexate therapy in patients with psoriasis [J]. *J Invest Dermatol.* 2008; 128(8): 1925–1929. [PubMed: 18256692]
- [50]. Obata H, Yahata T, Quan J, et al. Association between single nucleotide polymorphisms of drug resistance-associated genes and response to chemotherapy in advanced ovarian cancer[J]. *Anticancer Res.* 2006; 26(3B):2227–2232. [PubMed: 16821592]
- [51]. Siedlinski M, Boezen HM, Boer JM, et al. ABCC1 polymorphisms contribute to level and decline of lung function in two population-based cohorts [J]. *Pharmacogenet Genomics.* 2009; 19(9):675–684. [PubMed: 19687781]
- [52]. Pajic M, Murray J, Marshall GM, et al. ABCC1 G2012T single nucleotide polymorphism is associated with patient outcome in primary neuroblastoma and altered stability of the ABCC1 gene transcript[J]. *Pharmacogenet Genomics.* 2011; 21(5):270–279. [PubMed: 21317832]
- [53]. Zhao J, Yu BY, Wang DY, et al. Promoter polymorphism of MRP1 associated with reduced survival in hepatocellular carcinoma[J]. *World J Gastroenterol.* 2010; 16(48):6104–6110. [PubMed: 21182225]
- [54]. Innocenti F, Kroetz DL, Schuetz E, et al. Comprehensive pharmacogenetic analysis of irinotecan neutropenia and pharmacokinetics[J]. *J Clin Oncol.* 2009; 27(16):2604–2614. [PubMed: 19349540]
- [55]. Annemiek B, Corthals SL, Jongen LJ, et al. Mechanisms of peripheral neuropathy associated with bortezomib and vincristine in patients with newly diagnosed multiple myeloma: a prospective analysis of data from the hovan-65/GMMG-HD4 trial [J]. *Lancet Oncol.* 2010; 11(11):1057–1065. [PubMed: 20864405]
- [56]. Wojnowski L, Kulle B, Schirmer M, et al. NAD(P) H oxidase and multidrug resistance protein genetic polymorphisms are associated with doxorubicin-induced cardiotoxicity [J]. *Circulation.* 2005; 112(24):3754–3762. [PubMed: 16330681]
- [57]. Rajagopal A, Simon SM. Subcellular localization and activity of multidrug resistance proteins[J]. *Mol Biol Cell.* 2003; 14(8):3389–3399. [PubMed: 12925771]
- [58]. Wang H, Jin G, Wang H, et al. Genetic susceptibility of lung cancer associated with common variants in the 3'untranslated regions of the adenosine triphosphate-binding cassette B1 (ABCB1) and ABCC1 candidate transporter genes for carcinogen export[J]. *Cancer.* 2009; 115(3):595–607. [PubMed: 19107762]
- [59]. Yin JY, Han LF, Huang Q, et al. ABCC1 polymorphism Arg723Gln (2168G > A) is associated with lung cancer susceptibility in a Chinese population [J]. *Clin Exp Pharmacol Physiol.* 2011; 38(9):632–637. [PubMed: 21736601]
- [60]. Mafficini A, Ortombina M, Sermet-Gaudelius I, et al. Impact of polymorphism of multidrug resistance-associated protein 1 ABCC1 gene on the severity of cystic fibrosis [J]. *J Cyst Fibros.* 2011; 10(4):228–233. [PubMed: 21435954]
- [61]. Budulac SE, Postma DS, Hiemstra PS, et al. Multidrug resistance-associated protein-1 (MRP1) genetic variants, MRP1 protein levels and severity of COPD[J]. *Respir Res.* 2010; 11:60. [PubMed: 20487524]
- [62]. International HapMap Consortium. The international hapmap project[J]. *Nature.* 2003; 426(6968):789–796. [PubMed: 14685227]
- [63]. Pennisi E. Genomics. 1000 genomes project gives new map of genetic diversity[J]. *Science.* 2010; 330(6004):574–575. [PubMed: 21030618]
- [64]. Thomas PD, Campbell MJ, Kejariwal A, et al. PANTHER: a library of protein families and subfamilies indexed by function [J]. *Genome Res.* 2003; 13(9):2129–2141. [PubMed: 12952881]
- [65]. Ng PC, Henikoff S. Predicting deleterious amino acid substitutions[J]. *Genome Res.* 2001; 11(5): 863–874. [PubMed: 11337480]
- [66]. Pang GS, Wang J, Wang Z, et al. Predicting potentially functional SNPs in drug-response genes [J]. *Pharmacogenomics.* 2009; 10(4):639–653. [PubMed: 19374519]

- [67]. Wang Z, Wang J, Tantoso E, et al. Signatures of recent positive selection at the ATP-binding cassette drug transporter superfamily gene loci [J]. *Hum Mol Genet.* 2007; 16(11):1367–1380. [PubMed: 17412754]
- [68]. Bamshad M, Wooding SP. Signatures of natural selection in the human genome [J]. *Nat Rev Genet.* 2003; 4(2):99–111. [PubMed: 12560807]
- [69]. Mishra PJ, Humeniuk R, Longo-sorbello GS, et al. A miR-24 microRNA binding-site polymorphism in dihydrofolate reductase gene leads to methotrexate resistance [J]. *Proc Natl Acad Sci USA.* 2007; 104(33):13513–13518. [PubMed: 17686970]
- [70]. Hofmann MH, Bliedernicht JK, Klein K, et al. Aberrant splicing caused by single nucleotide polymorphism C516G > T [Q172H], a marker of CYP2B6 * 6, is responsible for decreased expression and activity of CYP2B6 in liver[J]. *J Pharmacol Exp Ther.* 2008; 325(1):284–292. [PubMed: 18171905]
- [71]. Sharom FJ. ABC multidrug transporters: structure, function and role in chemoresistance[J]. *Pharmacogenomics.* 2008; 9(1):105–127. [PubMed: 18154452]
- [72]. Nicolis E, Pasetto M, Cigana C, et al. The GCC repeat length in the 5'UTR of MRP1 gene is polymorphic: a functional characterization of its relevance for cystic fibrosis [J]. *BMC Med Genet.* 2006; 7:7. [PubMed: 16464259]
- [73]. Serajee FJ, Nabi R, Zhong H, et al. Polymorphisms in xeno-biotic metabolism genes and autism[J]. *J Child Neurol.* 2004; 19(6):413–417. [PubMed: 15446388]

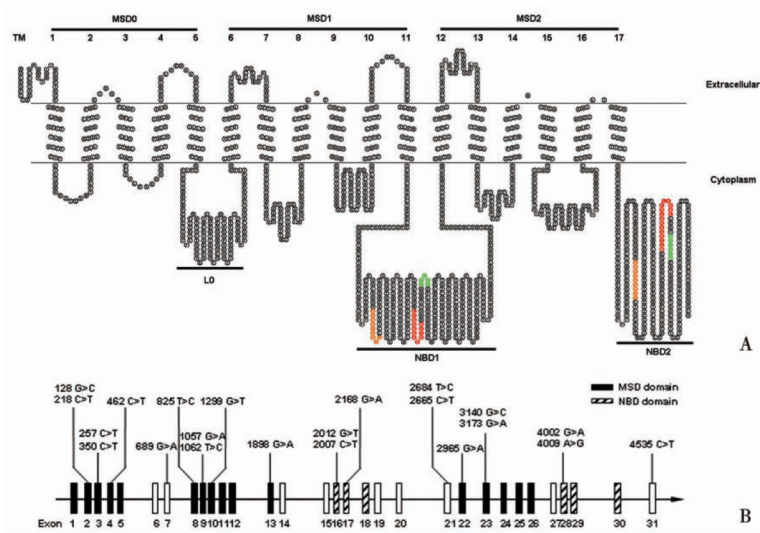


Fig. 1.
A:Schematic representation of the topological structure of MRP1/ABCC1 protein predicted using TOPO2 program with modification (<http://www.sacs.ucsf.edu/TOPO-run/wtopo.pl>). The consensus sequences of Walker A and B are highlighted in orange and green, respectively. The ABC-signature motif is highlighted in red. TM, transmembrane; MSD, membrane spanning domain; NBD, nucleotide-binding domain. B: Distribution of clinically relevant MRP1/ABCC1 exon polymorphisms.

Tab. 1

Clinically relevant substrates of MRP1/ABCC1 *

Type of substrates	Examples
Drugs	Anticancer drugs
	<i>Vinca</i> alkaloids: vinblastine and vincristine
	Epipodophyllotoxins: etoposide (VP-16) and teniposide
	Camptothecins: topotecan, irinotecan and SN-38
	Methotrexate and mitoxantrone
	Other drugs
	Anti HIV drugs; ritonavir and saquinavir
Heavy metal anions	Antibiotics: difloxacin and grepafloxacin
	Tyrosine kinase inhibitors: imatinib mesylate and gefitinib
	Arsenite
	Arsenate
	Trivalent and pentavalent antimonials
	Glutathione conjugates(-GS)
	Dinitrophenyl-GS
Glutathione conjugates(-GS)	Etacrynic acid-GS
	Doxorubicin-GS
	Cyclophosphamide-GS
	Melphalan-GS
	Aflatoxin B ₁ -epoxide-GS
	Hydroxynonenal-GS
	Prostaglandin A ₂ -GS
Glucuronide conjugates (-G)	Glutathione (GSH, GSSG)
	Bilirubin-G
	Estradiol 17βD-G
	Hyodeoxycholate-G
	Etoposide (VP-16)-G
	NS-38-G
	Sulfate conjugates (-S)
Sulfate conjugates (-S)	Estrone-3-S
	Taurocholate-3-S
	Dehydroepiandrosterone-3-S
	Sulfatolithocholyl taurine
Folates	Folic acid
	L-leucovorin
Toxicants	Aflatoxin B1
	Methoxychlor
	Fenitrothion
Others	Leukotrienes C4, D4 and E4
	Curcuminoids
	Calcein

* The primary references are available from the following reviews. [1, 10, 19, 71]

Tab. 2

Association of *MRP1/ABCC1* polymorphisms with therapeutic response

Polymorphisms	rs number	Amino acid exchange	Location	Drugs	Disease/Observation	References
G2012T	rs45511401	Gly671 Val	Exon 16	Atorvastatin	Hypercholesterolemia/No correlation	[42]
				Telatinib	Solid tumor/No correlation	[44]
				Induction Therapy	Leukemia/No correlation	[43]
G4002A	rs2239330	No change	Exon 28	Gemcitabine, cisplatin, taxane, methotrexate	Pancreatic cancer/No correlation	[45-47]
				Citalopram	Major depressive disorder/Strong correlation	[48]
G2168A	rs4148356	Arg723Gln	Exon 17	Platinum	Ovarian cancer/Correlation	[50]
				Taxane	Ovarian cancer/Correlation	[50]
A4009G	rs28364006	Ala1337Thr	Exon 28	Methotrexate	Psoriasis/Correlation	[49]
IVS23 G-1960A	rs2238476	No change	Intron 23	Methotrexate	Psoriasis/Correlation	[49]
IVS9 T-176C	rs35592	No change	Intron 9	Methotrexate	Psoriasis/Correlation	[49]
T2684C		No change	Exon 21		Leukemic/No correlation	[43]
C2007T	rs2301666	No change	Exon 16		Leukemic/No correlation	[43]
G2012T	rs45511401	Gly671 Val	Exon 16		Leukemic/No correlation	[43]
C2665T		No change	Exon 21		Leukemic/No correlation	[43]
IVS1 C-14840T	rs119774	No change	Intron 1	Montelukast	Asthma/Correlation	[28]
				Zileuton	Asthma/Correlation	[29]
GCC repeat		No change	5'UTR	Azithromycin	Cystic fibrosis/No correlation	[72]
IVS18 C -30G	rs2074087	No change	Intron 18	Taxanes	Ovarian cancer/No correlation	[47]

Tab. 3Association of *MRP1/ABCC1* polymorphisms with prognosis prediction

Polymorphisms	rs number	Amino acid exchange	Location	Disease/Observation	References
G2012T	rs45511401	Gly671Val	Exon 16	Neuroblastoma/Correlation	[52]
5'-UTR G-1666A	rs4148330	No change	5'UTR	Hepatocellular carcinoma/Correlation	[53]

Tab. 4

Association of *MRP1/ABCC1* polymorphisms with drug toxicity

Polymorphisms	rs number	Amino acid exchange	Location	Drugs	Drug toxicity/Disease/Observation	References
G4002A	rs2239330	No change	Exon 28	Irinotecan	Neutropenia/Solid tumor/Correlation	[54]
				Methotrexate	Overall MTX toxicity/Rheumatoid arthritis/No correlation	[46]
IVS11 -48C > T	rs3765129	No change	Intron 11	Irinotecan	Neutropenia/Solid tumor/Correlation	[54]
IVS9 A8G	rs35588	No change	Intron 9	Irinotecan	Neutropenia/Solid tumor/No correlation	[54]
T1684C	rs35605	No change	Exon 13	Irinotecan	Neutropenia/Solid tumor/No correlation	[54]
IVS30 A18G	rs212088	No change	Intron 30	Irinotecan	Neutropenia/Solid tumor/No correlation	[54]
IVS3 G-3198A	rs11075291	No change	Intron 3	Methotrexate	Hepatic and gastrointestinal toxicity/Psoriasis/Correlation	[49]
IVS4 G409A	rs1967120	No change	Intron 4	Methotrexate	Hepatic and gastrointestinal toxicity/Psoriasis/Correlation	[49]
IVS5 G413A	rs3784862	No change	Intron 5	Methotrexate	Hepatic and gastrointestinal toxicity/Psoriasis/Correlation	[49]
IVS5 A-7942G	rs246240	No change	Intron 5	Methotrexate	Hepatic and gastrointestinal toxicity/Psoriasis/Correlation	[49]
IVS5 G-1641A	rs3784864	No change	Intron 5	Methotrexate	Hepatic and gastrointestinal toxicity/Psoriasis/Correlation	[49]
IVS23 G-1960A	rs2238476	No change	Intron 23	Methotrexate	Hepatic and gastrointestinal toxicity/Psoriasis/Correlation	[49]
IVS14 C115T		No change	Intron 14	Methotrexate	Overall MTX toxicity/Rheumatoid arthritis/No correlation	[46]
IVS18 C-30G	rs2074087	No change	Intron 18	Methotrexate	Overall MTX toxicity/Rheumatoid arthritis/No correlation	[46]
G2012T	rs45511401	Gly671Val	Exon 16	Doxorubicin	Cardiotoxicity/NHL/Correlation	[56]
IVS16 A1695T	rs3887412	No change	Intron 16	Vincristine	Peripheral neuropathy/Multiple myeloma/Correlation	[55]

Tab. 5

Association of *MRP1/ABCC1* polymorphisms with disease susceptibility and severity

Polymorphisms	rs number	Amino acid exchange	Location	Diseases	Phenotype/Observation	References
3'-UTR T866A	rs212090	No change	3'-UTR	Lung cancer	Susceptibility/Correlation	[58]
				COPD	Severity/Correlation	[61]
3'-UTR C543T	rs3743527	No change	3'-UTR	Lung cancer	Susceptibility/No correlation	[58]
3'-UTR T1512C	rs212091	No change	3'-UTR	Lung cancer	Susceptibility/No correlation	[58]
T825C	rs246221	No change	Exon 8	Autism	Susceptibility/No correlation	[73]
G2168A	rs9448356	Arg723Gln	Exon 17	Lung cancer	Susceptibility/Correlation	[59]
5'-UTR G -260C	rs504348	No change	Promoter	Cystic fibrosis	Severity/Correlation	[60]
3'-UTR G3361A	rs4148382	No change	3'-UTR	COPD	Severity/Correlation	[61]
				Lung function	Severity/Correlation	[51]
5'-UTR C435G	rs504348	No change	5'-UTR	COPD	Severity/No correlation	[61]
IVS1 T5977G	rs4781699	No change	Intron 1	COPD	Severity/No correlation	[61]
IVS14 C-1575T	rs35621	No change	Intron 14	COPD	Severity/No correlation	[61]
				Lung function	Severity/Correlation	[51]
3'-UTR A2615G	rs212093	No change	3'-UTR	Lung function	Severity/Correlation	[51]